Mitochondrial phylogeography of European pond turtles (*Emys orbicularis*, *Emys trinacris*) – an update[∗]

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Abstract. Based on more than 1100 samples of *Emys orbicularis* and *E. trinacris*, data on mtDNA diversity and distribution of haplotypes are provided, including for the first time data for Armenia, Georgia, Iran, and the Volga, Ural and Turgay River Basins of Russia and Kazakhstan. Eight mitochondrial lineages comprising 51 individual haplotypes occur in *E. orbicularis*, a ninth lineage with five haplotypes corresponds to *E. trinacris*. A high diversity of distinct mtDNA lineages and haplotypes occurs in the south, in the regions where putative glacial refuges were located. More northerly parts of Europe and adjacent Asia, which were recolonized by *E. orbicularis* in the Holocene, display distinctly less variation; most refuges did not contribute to northern recolonizations. Also in certain southern European lineages a decrease of haplotype diversity is observed with increasing latitude, suggestive of Holocene range expansions on a smaller scale.

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The mitochondrial cytochrome *b* gene (cyt *b*) became a frequently used marker for inferring phylogeography in reptiles (e.g. Brown and Pestano, 1998; Burbrink et al., 2000; Carranza et al., 2000, 2002; Surget-Groba et al., 2001; Harris et al., 2002; Austin et al., 2003; Podnar et al., 2005; Poulakakis et al., 2005; Fritz et al., 2006a) and several studies on European pond turtles (*Emys orbicularis*, *Emys trinacris*) were based on this marker gene. While Lenk et al. (1999) provided a nearly rangewide phylogeography, their study suffered from a patchy sampling for many parts of the range

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(North Africa, Iberian and Apennine Peninsulas, France, East Europe, Kazakhstan, Turkey, Caucasus, Iran and Turkmenistan). Subsequent papers used a much denser sampling but covered only small regions (Fritz et al., 2004, 2005a, b, 2006b; Kotenko et al., 2005). Here we provide a range-wide update on haplotype diversity and distribution, and present for the first time data on pond turtles from Armenia, Georgia, Iran, and the Volga, Ural and Turgay River Basins of Russia and Kazakhstan. This note is based on more than 1100 cyt *b* sequences of known-locality samples and specimens of unknown geographic origin and is likely to represent the largest data set ever published for western Palaearctic vertebrates. Our paper is aimed as a starting-point for further research, indicating still badly-sampled regions and suggesting future research directions.

Sampling techniques, PCR and sequencing are described in Lenk et al. (1999) and Fritz et al. (2005a). Besides samples from native wild-caught turtles or known-locality specimens from captive breeding projects, samples of confiscated, pet trade, or wild-caught allochthonous turtles were studied. Sequences were approximately 970-1125 bp long and aligned manually for haplotype determination. Haplotype nomenclature follows Lenk et al. (1999) and Fritz et al. (2004, 2005a, b, 2006b): Roman numerals designate major clades of haplotypes as revealed by phylogenetic analyses (=mtDNA lineages); within each lineage individual haplotypes are distinguished by consecutive letters.

In addition to the previously identified 48 haplotypes in nine lineages (I-IX; Lenk et al., 1999; Fritz et al., 2004, 2005a, b, 2006b) we found eight new haplotypes, belonging to already known lineages. For many haplotypes described in earlier studies, distributional data are provided here for the first time. The geographic origin of lineage IX (with haplotype IXa only), discovered in a pet trade turtle (Fritz et al., 2004), is still unknown however (table 1).

Based on an alignment of 1031 bp, relationships of haplotypes are illustrated using a TCS 1.21 parsimony network (Clement et al., 2000) and a MP strict consensus tree rooted with the closely related Nearctic taxa *Actinemys marmorata* and *Emydoidea blandingii* (PAUP* 4.0b10, TBR branch-swapping algorithm, all characters with equal weight, stepwise random addition of 10 sequences; Swofford, 2002). For the ingroup taxa 957 characters were constant, 30 were variable but parsimony-uninformative and 44 were parsimony-informative; for all taxa 877 characters were constant, 33 variable characters were parsimony-uninformative and 121 were parsimony-informative. Maximum likelihood estimates for genetic differences of all nine lineages and most haplotypes were depicted in a recently published ML phylogram (Fritz et al., 2005a); uncorrected *p* distances were reported in the same study.

The network (fig. 1) is flock-like and haplotypes cluster into eight major branches. One branch splits into lineages I and II (*E. o. orbicularis* and related subspecies); the other eight branches correspond with lineages III (*E. trinacris*) and lineages IV to IX (remaining *E. orbicularis* subspecies). Lineages I and II are interconnected over two loops; a further loop occurs within lineage IV. Under MP phylogenetic analysis (fig. 2), monophyly of haplotypes of lineages II, III, IV, V and VII is well-supported, while monophyly for lineage I and VI haplotypes is only weakly supported. Lineages VIII and IX are represented by only one haplotype each that is located outside of all other lineages. Lineage III (*E. trinacris*) constitutes the sister group of a weakly supported clade containing lineages I-II and IV-IX (*E. orbicularis*); a wellsupported clade of lineages I and II is the sister group of the remaining lineages IV-IX within *E. orbicularis*. Bootstrap support for monophyly of the clade comprising lineages IV-IX is below 50% however. Within lineage VI, North African haplotypes (VIc, VIf) are paraphyletic with respect to European haplotypes (VIa, VIb, VId, VIe).

Regarding geographic distribution and phylogeography, our new data confirm the findings of

Figure 1. Parsimony network (TCS, spring tree) of all 56 known mtDNA haplotypes of *Emys orbicularis* (haplotypes of lineages I-II, IV-IX) and *Emys trinacris* (haplotypes of lineage III). Branch lengths correspond to inferred number of nucleotide changes along each branch; open circles and dots, identified or hypothesized haplotypes, respectively. Each line between circles or dots indicates one substitution; circle size corresponds to approximate frequency of haplotypes (table 1); size classes: 1, 2-9, 10-25, 26-50, 51-150, 151-200, *>*200 recorded haplotypes.

Lenk et al. (1999) and Fritz et al. (2005a) in that a high diversity of distinct mtDNA lineages and haplotypes occurs in southern regions where putative glacial refuges were located (fig. 3, table 1). With the advent of Holocene warming, more northerly parts of Europe and adjacent Asia were rapidly recolonized by *E. orbicularis* from few refuges in the Balkans and the northern Black Sea/northern Caucasus region, resulting in the respective mtDNA lineages in decreasing haplotype diversity with increasing distance to the refuge (long distance dispersal model of Hewitt, 1996); most refuges did not contribute to northern recolonizations.

With exception of a doubtful record of haplotype VIIa, the northernmost parts of the range of *E. orbicularis* are occupied only by three haplotypes (Ia, IIa and IIb), while lineages I and II exhibit a much higher diversity in the south. The same pattern, suggestive of Holocene range

Figure 2. Strict consensus of six equally parsimonious trees of all *Emys* haplotypes using sequences of *Actinemys marmorata* (accession numbers AJ131430, U81344) and *Emydoidea blandingii* (AF258869, AJ131432) as outgroup $(CI = 0.7585, RI = 0.9120, RC = 0.6917; 207 steps)$. Numbers above nodes are bootstrap values (1000 resamplings) greater than 50. The six equally parsimonious trees differ only in the branching pattern of lineage IV haplotypes.

expansions on a smaller scale, is observed in lineages IV and V with several endemic haplotypes in the southernmost parts of their ranges; northern portions harbour only haplotypes IVa or Va. A previous record of haplotype VIIa for the northern Caspian Sea region (Lenk et al., 1999) should be treated with care. It was based on a turtle kept in the zoological garden of Almaty (Kazakhstan), and we cannot exclude lo-

Figure 3. Geographic distribution of mtDNA lineages in *Emys orbicularis* (I-II, IV-IX) and *Emys trinacris* (III). Neighbouring localities combined; overlapping symbols indicate syntopic occurrence of distinct lineages; question marks, doubtful records for lineage III in Calabria (southern Italy) and lineage VII in the northern Caspian region. Evidently allochthonous specimens not considered.

cality confusion. Otherwise, lineage VII occurs far away, in the eastern central Caucasus and along the south coast of the Caspian Sea (fig. 3, table 1).

The wide distribution of lineage I haplotypes around the Black Sea suggests that the entire Black Sea region served as glacial refuge. On the other hand, the localized distribution of endemic lineage I haplotypes (table 1) indicates that distinct microrefuges existed there. An unexpected finding was haplotype Ia in eastern Georgia (regions of Gori and Tbilisi: one record each from Kareli, Gldanula River and Udabno). These sites are situated in the Kura River system that drains into the Caspian Sea. In the same river system occurs another mitochondrial lineage (VII) farther east, suggesting that lineages I and VII meet somewhere in the central Caucasus. This situation is echoed in the phylogeography of *Testudo graeca*. Also in this species two distinct mitochondrial lineages meet or occur in

closest neighbourhood in the central Caucasus (Fritz et al., 2007).

Besides the wide-spread haplotype IIa, the most differentiated haplotype of lineage II (IIf) occurs on the south-eastern Balkan Peninsula (table 1), suggesting the refuge for lineage II was located there. From the southeastern Balkans, lineage II most probably spread along the valleys of the Vardar and Južna Morava Rivers northward to the Danube catchment basin from where central and western Europe (Fritz et al., 2005b) and, via the Moravian Gate, the Oder River Basin (Germany, Poland) were reached.

As for lineage IV, displaying a classic circum-Adriatic distribution, two distinct refuges existed in southern Italy and the southernmost Balkan Peninsula. The lack of the common haplotype IVa, distributed in Italy, Istria, Dalmatia, Corfu and Evvia, and the occurrence of endemic haplotypes in the Peloponnesus and on

Cephalonia (IVb, IVc, IVg) provide evidence for the colonization of the west coast of the Balkans, Corfu and Evvia from southern Italy and not from the geographically closer southern Balkanic refuge (Fritz et al., 2005a).

Like in *Mauremys leprosa* (Fritz et al., 2006a), mountain chains constitute major biogeographic barriers for mtDNA lineages in *E. orbicularis* (fig. 3). The Pyrenees separate the Ibero-Maghrebinian lineage VI from lineages II and V in the north, and the Apennines separate lineages IV and V in Italy. In the Balkan Peninsula, the Dinarid and Pindos Mts. are a barrier between lineage IV along the west coast and lineages I and II in the east, and south of the Alps occur lineages IV and V while in the north lineage II is distributed. In East Europe the Carpathians act as barrier between lineage II in the south and lineage I in the north; farther eastward the Greater Caucasus separates lineages I and VII and the Taurus Mts. (Turkey) divide lineages I and VIII. Syntopic occurrences of distinct mtDNA lineages, most probably as result of Holocene range expansions, are confined to narrow contact zones in close proximity of these mountain chains in the northern Iberian Peninsula, France and the southern Apennine and Balkan Peninsulas. Obviously, marshlands along sea coasts and river courses were used as colonization corridors during range expansions, enabling development of contact zones.

The basal position of the North African haplotypes VIc and VIf in the MP tree and in the network branch of lineage VI (figs 1, 2) suggests that the Iberian haplotypes VIa, VIb, VId and VIe are derived from North African haplotypes. This implies a complicated colonization history, from Europe to North Africa and back to Europe, as recently proposed for the ribbed newt *Pleurodeles waltl* (Veith et al., 2004). Denser sampling is still needed in North Africa however, especially in Algeria and Tunisia. Some Moroccan amphibians and reptiles that occur in habitats resembling those of *E. orbicularis* (*Pleurodeles waltl*, *Mauremys l. leprosa*) are replaced in eastern Algeria and Tunisia by other taxa (*P. nebulosus*, *P. poireti*, *M. l. saharica*, Carranza and Arnold, 2003; Carranza and Wade, 2004; Veith et al., 2004; Fritz et al., 2006a; distinct mitochondrial lineages of *Natrix maura*, Guicking et al., 2006), suggestive of a similar pattern in *E. orbicularis*. Further sampling is also needed in Turkey, Turkmenistan, in the Caucasus and in the northern Black Sea region to reveal haplotype diversity and potential contact zones there; one of these regions must harbour lineage IX.

Besides mtDNA haplotyping, future research should focus on gene flow along contact zones of distinct mitochondrial lineages. A promising approach will be using microsatellites as markers.

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